

Taking a Gene to the Bench

How do we test a gene's actual job at the bench?

CARRY FORWARD

Many tests inflate false positives, so thresholds and independent replication must be planned before results are seen.

10-YEAR TAKEAWAY

Orthogonal assays answer different mechanism questions: where RNA or protein is and what changes after perturbation.

Cells differentiate	Development sculpts	Cells move	Development can go wrong
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1. Observe the analogy before naming the biology

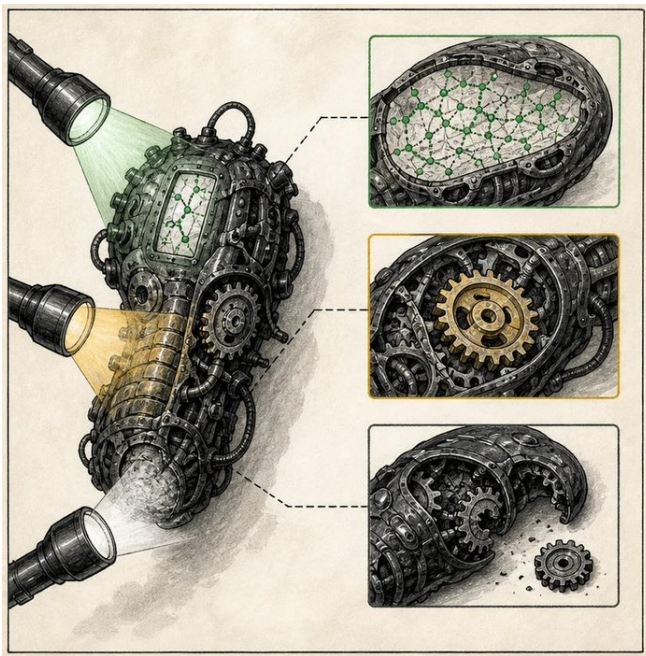


Illustration: Three flashlights reveal message, material, and system failure

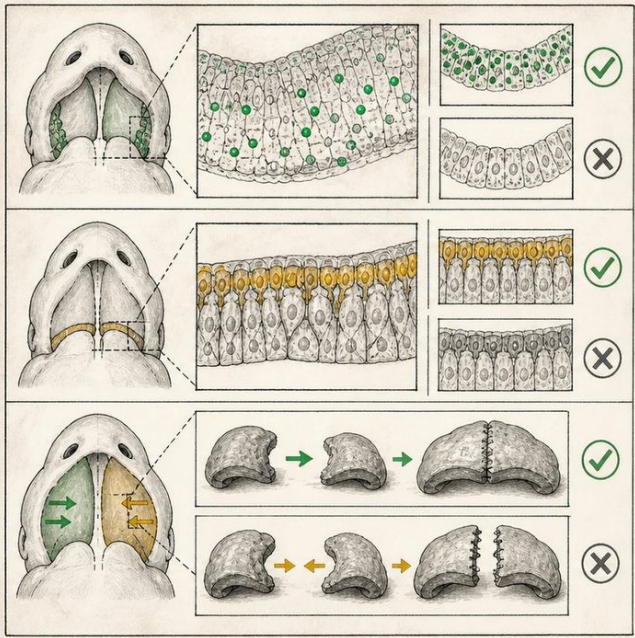
- 1. Which light shows where the message is?
- 2. Which shows where the working part is?
- 3. Which reveals what changes when the part is missing?

2. Turn observations into rules

- 1. Rule 1: Match each assay to one question. Explain it in your own words.
- 2. Rule 2: Include negative and positive controls. Explain it in your own words.
- 3. Rule 3: Combine location evidence with perturbation evidence before claiming function. Explain it in your own words.

Where the analogy stops: Assays produce measured signals with noise and controls, not perfectly revealing beams.

3. Map the rules onto the biomedical example



Biomedical teaching illustration: Test a candidate developmental gene with three assays

Analogy step	Biology step	How the relationship stays the same
Wiring-map light	RNA in situ hybridization	_____
Working-part light	Protein immunohistochemistry	_____
Removal test	Knockout or knockdown phenotype	_____

EXPERIMENTAL DESIGN LESSON 9 | CASE FILE AND DECISION

4. Use the case evidence

ID	Finding supplied in the case	Why it matters
EXP09-E1	RNA signal appears in developing palatal shelves at the relevant stage.	The gene is transcribed in a useful place and time.
EXP09-E2	Protein staining appears in palatal epithelium and is absent in the no-primary-antibody control.	The protein location is supported by a specificity control.
EXP09-E3	Perturbed embryos show altered shelf elevation more often than matched controls.	Changing the gene is associated with a phenotype, but other effects still need checks.

5. Make the clinical decision

You are planning the bench follow-up to a replicated locus. The team has funding for three assays and must decide whether one expression image alone proves function.

- A. Combine RNA location, protein location, perturbation, and controls.
- B. Use one bright stain as proof the gene causes cleft palate.
- C. Skip controls because the candidate came from GWAS.

Decision. Choose the assay set and state the specific question each assay answers.

Claim ceiling: You may connect expression and perturbation evidence. You may not treat location alone as causal proof.

Claim. State the choice you can defend.

Evidence. Use these labeled IDs: EXP09-E1 + EXP09-E2.

Reasoning. Explain why the evidence supports the claim and stays inside the claim ceiling.

6. Reduce the lesson to one lasting idea

Take-home. Orthogonal assays answer different mechanism questions: where RNA or protein is and what changes after perturbation.

1. What does in situ hybridization locate?
2. Why is expression alone not proof of function?

Glossary

Term	Plain-English definition
knockout	An organism or cell engineered to have a specific gene switched off, used to reveal what that gene normally does.
conditional knockout	A genetic tool that switches a gene off only in certain cells or at a chosen time, so researchers can study its role without affecting the whole animal.
in situ hybridization	A lab method that uses a labeled probe to show exactly where a specific gene's RNA or DNA sits inside a tissue or cell.
immunohistochemistry	A lab technique that uses antibodies tagged with stain to show exactly where a specific protein sits within a tissue slice.
penetrance	The fraction of people carrying a disease-linked genotype who actually show the trait.
control	The comparison condition that isolates the effect of the variable being tested by keeping everything else the same.

Next lesson unlock

A knockout removes a gene and a stain shows where it sits, but both are slow and blunt; building a conditional-knockout mouse takes many crosses and many months. Is there a faster, more precise way to edit a gene on purpose and read out exactly what it does?

The journal links on the lesson page are optional. Everything required for this guided-notes set is already supplied here and in the case file.